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Functional principal component analysis and sparse-group LASSO to identify associations between biomarker trajectories and mortality among hospitalized SARS-CoV-2 infected individuals

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Abstract

Background A substantial body of clinical research involving individuals infected with severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) has evaluated the association between in-hospital biomarkers and severe SARS-CoV-2 outcomes, including intubation and death. However, most existing studies considered each of multiple biomarkers independently and focused analysis on baseline or peak values.

Methods We propose a two-stage analytic strategy combining functional principal component analysis (FPCA) and sparse-group LASSO (SGL) to characterize associations between biomarkers and 30-day mortality rates. Unlike prior reports, our proposed approach leverages: 1) time-varying biomarker trajectories, 2) multiple biomarkers simultaneously, and 3) the pathophysiological grouping of these biomarkers. We apply this method to a retrospective cohort of 12, 941 patients hospitalized at Massachusetts General Hospital or Brigham and Women's Hospital and conduct simulation studies to assess performance.

Results Renal, inflammatory, and cardio-thrombotic biomarkers were associated with 30-day mortality rates among hospitalized SARS-CoV-2 patients. Sex-stratified analysis revealed that hematogolical biomarkers were associated with higher mortality in men while this association was not identified in women. In simulation studies, our proposed method maintained high true positive rates and outperformed alternative approaches using baseline or peak values only with respect to false positive rates.

Conclusions The proposed two-stage approach is a robust strategy for identifying biomarkers that associate with disease severity among SARS-CoV-2-infected individuals. By leveraging information on multiple, grouped biomarkers' longitudinal trajectories, our method offers an important first step in unraveling disease etiology and defining meaningful risk strata.

Keywords SARS-CoV-2, Biomarkers, Sparse group LASSO, Functional data analysis, Functional principal component analysis

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Background

Since the outbreak of severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) in December 2019, more than 670 million confirmed cases and 6.8 million associated deaths have been reported worldwide, with a large proportion of these deaths preceded by hospitalization [1]. The vast amount of data collected and stored in electronic health records among hospitalized patients provides an opportunity to identify early predictors of severe disease. Ultimately understanding the relationship between patient level in-hospital data, including early biomarker trajectories, and severe outcomes may inform disease etiology, risk stratification, and resource allocation.

Among SARS-CoV-2 infected individuals, multiple biomarkers are typically measured repeatedly over the duration of hospitalization. An extensive literature has identified correlations between biomarker levels and severe outcomes, including intubation, admission to intensive care units, and death among hospitalized SARS-CoV-2 infected individuals [2–14]. However, most existing studies considered each biomarker independently [2-9] with a few exceptions that applied penalized regression and other machine learning techniques [10, 11]. None of these manuscripts, to our knowledge, accounted for the pathophysiological relationships among biomarkers. Moreover, repeatedly measured biomarkers were typically reduced to baseline or peak values [2-8, 10, 11], with again a small number of exceptions, including one report using linear mixed-effects models to account for the entire biomarker trajectory [9]. To our knowledge, analyses that simultaneously consider multiple biomarkers as well as their longitudinal trajectories in evaluating associations with severe SARS-CoV-2 outcomes have not been reported.

Methods for joint modeling of multiple longitudinal biomarkers and time-to-event outcomes have also been described [15–18]. However, as these methods generally require computationally intensive procedures, such as multi-dimensional numerical integration or complex Bayesian sampling schemes, approaches incorporating variable selection among multiple biomarkers into the joint modeling framework remain limited [18, 19]. A scalable alternative involves application of multivariate functional principal component analysis (FPCA) [20] to reduce each biomarker trajectory to a set of scores and then using these scores as covariates in a survival model [21-23]. Application of FPCA and survival modeling has been limited to prediction of the time-to-event outcome. To allow for variable selection in this context, we propose a two-stage analytic strategy that combines FPCA and sparse-group LASSO (SGL) [24], abbreviated as FPCA-SGL, to characterize associations between multiple biomarker trajectories and mortality, while also leveraging the pathophysiological grouping of these biomarkers.

Methods

Study population

Data derived from a retrospective cohort of 12,941 patients infected with SARS-CoV-2 based on hospital record ICD-10 codes (U07.1, B34,2, and B97.29) and positive PCR tests between March 1, 2020 and November 30, 2021 were used for analysis (Table 1). All patients were hospitalized at Massachusetts General Hospital or Brigham and Women's Hospital (MGB) within 5 days prior to and 30 days after a positive SARS-CoV-2 test. Patients hospitalized for less than 24 hours or with an unknown duration were excluded.

Data pre-processing

The primary outcome is 30-day mortality since hospital admittance. Death records from both the MGB Enterprise Data Warehouse (EDW) and the Massachusetts Registry of Vital Records and Statistics were obtained, and in the case of an inconsistency between the two sources, death dates from the Registry were adopted. Exposures are repeated laboratory measurements of m = 20 routine biomarkers collected during hospitalization up to 30 days. Biomarker data were extracted from MGB EDW and if there were multiple measurements of a biomarker for one patient within a 24-hour period, the mean value of the measurements was used. Censored laboratory measurements were treated as known at the cut-off value. Demographic information including age, sex, race/ethnicity and body mass index (BMI) was obtained from MGB EDW. Biomarkers were divided into six categories based on their pathophysiological functions, as shown in Supplementary Table 1.

Statistical analysis

A two-stage analytic approach was considered. First, FPCA was performed separately on each of the $k = 1, 2, \dots, K$ biomarkers. Each biomarker's repeated measurements were treated as functional data, i.e., independent realizations of a smooth random function $X_k(t)$ [20]. Through spectral decomposition of the covariance operator, FPCA reduces the functional data into eigenfunctions $\phi_{km}(t)$ for $m = 1, \dots, M$, referred to as functional principal components (FPCs). Each individual *i* has a set of coefficients for these eigenfunctions called FPC scores, denoted as A_{kim} . Thus the trajectory of one biomarker for patient *i*, $X_{ki}(t)$, can be expanded as

$$X_{ki}(t) = \mu_k(t) + \sum_{m=1}^{\infty} A_{kim} \phi_{km}(t) \approx \mu_k(t) + \sum_{m=1}^{M} A_{kim} \phi_{km}(t),$$

where $\mu_k(t) = \mathbb{E}[X_k(t)]$ is the mean function. Then each patient's FPC scores, $A_{kim}, m = 1, \dots, M$, characterize

Table 1 Demographic characteristics, survival outcomes and time in hospitalization stratified by sex

	Female (n=6300)	Male (n=6641)	Total (n=12941)
Age			
Age <u><</u> 50	2138 (33.9%)	2007 (30.2%)	4145 (32.0%)
50 <age<u><65</age<u>	1530 (24.3%)	2061 (31.0%)	3591 (27.7%)
Age>65	2632 (41.8%)	2573 (38.7%)	5205 (40.2%)
Race			
White, non-Hispanic	3610 (57.3%)	3961 (59.6%)	7571 (58.5%)
Black, non-Hispanic	820 (13.0%)	698 (10.5%)	1518 (11.7%)
Asian, non-Hispanic	213 (3.4%)	251 (3.8%)	464 (3.6%)
Hispanic	1317 (20.9%)	1341 (20.2%)	2658 (20.5%)
Other/Unknown	340 (5.4%)	390 (5.9%)	730 (5.6%)
BMI			
Normal (BMI<25)	1368 (21.7%)	1341 (20.2%)	2709 (20.9%)
Overweight (25 <u><</u> BMI<30)	1528 (24.3%)	2050 (30.9%)	3578 (27.6%)
Obese (BMI≥30)	2502 (39.7%)	2132 (32.1%)	4634 (35.8%)
Missing	902 (14.3%)	1118 (16.8%)	2020 (15.6%)
Deaths			
Number of deaths	488 (7.7%)	710 (10.7%)	1198 (9.3%)
Time (days) to death	10.5 (5, 16)	11 (6, 18)	11 (6, 17)
Time (days) in hospital			
Overall	5 (3, 9)	5 (3, 10)	5 (3, 10)
For dead within 30 days	8 (4, 13)	8 (5, 15)	8 (4, 14)
For alive and discharged within 30 days	5 (3, 8)	5 (3, 8)	5 (3, 8)
For alive and still in hospital after 30 days	38.5 (33, 46.25)	41 (34, 54)	39.5 (34, 52.5)

Summary measures are presented as count (percentage) for categorical variables and median (interquartile range) for continuous variables

the variation of individual level biomarker trajectories from the sample mean function. We adopted the PACE method which computes the FPC scores as conditional expectations because it is suitable for sparse and irregularly spaced longitudinal data like our biomarker data [25]. To implement FPCA using the PACE approach, we used the fdapace package in R [26]. Based on the cumulative percentage of variance explained, we determined the number of FPCs (*M*) to adopt, resulting in $K \times M$ exposure variables. Missing FPC scores were imputed using MICE based on the FPC scores of all other biomarkers [27].

Second, using the SGL package in R, we performed Cox SGL regression with the $K \times M$ FPC scores as the exposure variables, while adjusting for *J* demographic characteristics Y_{ij} including race/ethnicity, age (indicator for > 50 years), and BMI (orthogonal polynomials of degree 2):

$$h_i(t) = h_0(t) \exp\left[\left(\sum_{k=1}^K \sum_{m=1}^M \delta_{km} A_{kim}\right) + \sum_{j=1}^J \zeta_j Y_{ij}\right],$$

where there are $K \times M + J$ regression parameters to be estimated, collectively denoted as $\boldsymbol{\beta} = \{\delta_{km}, \zeta_j\}$ for $k = 1, \dots, K, m = 1, \dots, M$, and $j = 1, \dots, J$.

SGL estimates the β 's using a weighted combination (controlled by a hyperparameter η) of group LASSO l_1 -penalty term and the standard parameter-wise LASSO l_1 -penalty term to induce both groupwise and within-group sparsity [24]:

$$(1-\eta)\lambda\sum_{l=1}^{L}\sqrt{p_l}\left\|\beta^{(l)}\right\|_2+\eta\lambda\|\beta\|_1$$

where the grouping $l = 1, \dots, L$ represents biomarkers' six pathophysiological categories and one group for all demographic characteristics, i.e. for $l = 1, 2, \dots, 6$, $\beta^{(l)} = \{\delta_{km}\}$ for k over all biomarkers in that pathophysiological group and $m = 1, \dots, M$ for each biomarker's M FPC scores; and $\beta^{(7)} = \{\zeta_j\}$ for $j = 1, \dots, J$.

This two-stage strategy, FPCA decomposition of the biomarker trajectoris followed by SGL, allowed us to identify biomarkers associated with 30-day mortality while accounting for within-group correlations between biomarkers as well as the time varying biomarker trajectories. Both stages of analysis were stratified by sex.

We selected the overall regularization parameter λ through a 10-fold cross validation (CV) from a pre-specified sequence of 100 λ values. The sequence of 100 candidate λ values were chosen such that the maximum, λ_{max} , was the smallest possible λ that shrunk all coefficients to zero, the minimum, λ_{min} , was set equal to $\lambda_{max}/100$, and all other λ values were spaced equally between λ_{min} and λ_{max} . Eventually, we selected λ_{1se} which was the largest value of λ such that the CV error, defined as the CV negative log likelihood, was within 1 standard error of the minimum.

We set the weight for the group LASSO and LASSO penalty terms to $1 - \eta = 0.7$ and $\eta = 0.3$, respectively to get a group LASSO structure with limited within-group sparsity. This reflects that biomarkers are expected to exhibit group structure due to their underlying pathophysiological relationships, while still allowing individual biomarkers or FPC scores to be excluded from the model to enhance sparsity. Alternatively, this weight parameter η could be tuned via an additional layer of CV. As a sensitivity analysis, we fit the SGL model with different weights, $\eta = 0.05, 0.50, 0.70, 0.95$ respectively, to examine whether the results were sensitive to the choice of this hyperparameter.

For comparison, we considered application of SGL using only the baseline or peak measurements of each of the *K* biomarkers in place of the FPC scores. Here we log transformed the baseline and peak measurements to ensure they were approximately normally distributed, and again imputed missing values with available biomarker measures using the MICE package in R [27]. Analyses were again stratified by sex and adjusted for race/ethnicity, age, and BMI while accounting for the pathophysiological groupings of biomarkers.

Simulation studies

To characterize the performance of our proposed twostage approach, we conducted simulation studies including 200 repetitions with sample sizes of n = 2000 for each condition [21, 28] (Supplementary Methods). We first simulated trajectories of four biomarkers belonging to two groups, denoted as $Z_{ki}(t)$ where k = 1, 2, 3, 4 and $i = 1, 2, \dots, n$. Biomarkers k = 1, 2 were in the first group with relatively low within-group correlation, and biomarkers k = 3, 4 were in the second group with relatively high within-group correlation. $Z_{ki}(t)$ were simulated under three models: Model 1 was a linear mixed-effects model (LME) with a linear time trend; Model 2 was a LME with a quadratic term for time; and Model 3 was a LME with a 3-knot spline function for time.

Death times were simulated based on $Z_{ki}(t)$ using inverse transform sampling on the survival function derived from the following hazard function: $h_i(t) = h_0(t) \exp[\alpha_1 \times Z_{1i}(t) + \alpha_2 \times Z_{2i}(t) + \alpha_3 \times Z_{3i}(t)]$ $+\alpha_4 \times Z_{4i}(t)$]. The association parameters $\alpha_1, \alpha_2, \alpha_3, \alpha_4$, were specified based on four scenarios: Scenario 1, only the low correlation biomarker group was associated with mortality; Scenario 2, only the high correlation biomarker group was associated with mortality; Scenario 3, both biomarker groups were associated with mortality; Scenario 4, neither of the two biomarker groups was associated with mortality (Table 2). Measurement error was added to the true trajectories to simulate observed trajectories. Lastly, we simulated censoring by truncating observed trajectories at the patient's time of death or discharge (Supplementary Methods).

For the primary analysis we applied FPCA-SGL using the top three FPC scores (M = 3) for each biomarker as the exposure variables, i.e. $K \times M = 4 \times 3 = 12$ variables. The weights for the group LASSO and LASSO penalty terms were set to $1 - \eta = 0.95$ and $\eta = 0.05$, respectively because our simulated scenarios were a group LASSO case. To assess performance of our proposed approach, we reported the true positive rate (TPR), defined as the proportion of simulations where truly non-zero coefficients were selected, and the false positive rate (FPR), defined as the proportion of

Table 2 Set up of simulation studies

	α1	α2	α3	α4
Model 1 (LME with a linear time trend)				
Scenario 1 (low correlation)	1	1	0	0
Scenario 2 (high correlation)	0	0	1	1
Scenario 3 (both groups)	1	1	1	1
Scenario 4 (null case)	0	0	0	0
Scenario 5ª (complete null case)	0	0	0	0
Model 2 (LME with a quadratic term for	time)			
Scenario 1 (low correlation)	1	1	0	0
Scenario 2 (high correlation)	0	0	1	1
Scenario 3 (both groups)	0.5	0.5	0.5	0.5
Scenario 4 (null case)	0	0	0	0
Scenario 5ª (complete null case)	0	0	0	0
Model 3 (LME with a 3-knot spline func	tion for ti	me)		
Scenario 1 (low correlation)	1	1	0	0
Scenario 2 (high correlation)	0	0	1	1
Scenario 3 (both groups)	1	1	1	1
Scenario 4 (null case)	0	0	0	0
Scenario 5ª (complete null case)	0	0	0	0

^a Scenario 5 is an additional scenario based on Scenario 4 where we did not censor biomarker trajectories by death times. Details about this scenario were explained in the Results section

simulations where truly zero coefficients were selected. We also compared FPCA-SGL to two simpler comparator approaches using baseline or peak measurements alone.

Results

Application using MGB cohort

The MGB cohort was composed of 12,941 patients, 32% were \leq 50 years of age, 20.5% Hispanic, 11.7% Black/non-Hispanic, and 35.8% obese (BMI \geq 30) (Table 1). 1,198 patients (9.3%) died within 30-days of hospitalization, with a higher proportion in males than in females (10.7% vs. 7.7%) (Table 1). Supplementary Table 1 summarizes laboratory measurements for 20 biomarkers. More than 60% of patients had at least one measurement on each of the 20 biomarkers, with a median number of measurements for each biomarker ranging from 1 to 6, except that d-dimer had fewer measurements.

We performed FPCA on each of the 20 biomarkers, stratified by sex. Supplementary Fig. 1 displayed the mean function and corresponding FPCs of each biomarker. Across the 20 biomarkers, the first three FPCs cumulatively explained a median of 97.39% [IQR = (95.32%, 99.22%)] and 97.49% [IQR = (96.06%, 98.48%)] of the total variance among females and males, respectively. Therefore, we picked M = 3 FPCs for each of the M biomarkers. FPC scores were approximately normally distributed (Supplementary Fig. 2).

To better illustrate how each patient's M = 3 FPC scores could represent the variation of their individual biomarker trajectories from the mean function, Supplementary Fig. 3 plotted the trajectories of blood urea nitrogen (bun) of three male patients with different FPC scores $A_{k=1,i=\{1,2,3\},m=\{1,2,3\}}$. It shows how each individual's trajectory decomposes into a linear combination of the mean function and three eigenfunctions, resulting in different individual-specific FPC scores.

The pairwise baseline biomarker correlations were similar among females and males (Fig. 1A, B). The renal, hematological, and the hepatic groups exhibited high within-group correlation while the cardio-thrombotic, inflammatory and metabolic groups presented low within-group correlation. The across-group correlations were generally low (Fig. 1A, B). The pairwise peak biomarker correlations showed similar patterns (Fig. 1C, D). Biomarker peak and baseline values were approximately normally distributed after log transformation, imputation and standardization, with the exception of estimated glomerular filtration rate and total bilirubin (Supplementary Fig. 4).

Using the FPCA-SGL approach with $K \times M = 20 \times 3 = 60$ FPC scores as exposure variables, we found biomarkers in the renal and inflammatory groups to be strongly associated with mortality in both males and females. In the cardio-thrombotic group, only d-dimer appeared to be associated with mortality. Biomarkers in the hepatic groups showed slight associations while the metabolic group was not associated with mortality. The hematological group was associated with mortality among males but not females (Fig. 2).

For comparison, we used each of the 20 baseline measurements and the 20 peak measurements as exposure variables, and fitted SGL Cox regressions stratified by sex. Using baseline measurement, all biomarker groups except for the hematological group among females and the metabolic group among males were associated with mortality, with some degree of within-group sparsity observed (Fig. 3). Using peak measurements, most biomarkers across all groups except for the metabolic group were associated with mortality, with almost no withingroup sparsity (Fig. 3). Results from our simulation studies, as presented below, demonstrated that using baseline or peak measurements can result in high false positive rates.

As a sensitivity analysis, we applied the FPCA-SGL approach with the same 60 FPC scores while changing the weight hyperparameter η , with larger η implying more LASSO than group LASSO structure. Supplementary Fig. 5 displayed four similar heatmaps as Fig. 2 for four different weight values: $\eta = 0.05, 0.50, 0.70, 0.95$. The associations were similar with the main analysis: the renal and inflammatory groups were still strongly associated with mortality in both sexes; in the cardio-thrombotic group, d-dimer showed associations and creatine phosphokinase only showed trivial associations when a group LASSO structure was enforced under $\eta < 0.3$; the hepatic group was slightly associated with mortality and under small η the entire group was not selected, while larger η revealed that the aspartate aminotransferase and albumin in the group were the biomarkers driving these associations; the metabolic group was not associated with mortality; the hematological group showed associations still only among males but not females. Therefore, this analysis demonstrated that the results were not sensitive to the choice of the weight hyperparameter.

Simulation studies

Both our proposed two-stage FPCA-SGL method and the simpler comparator methods using baseline or peak measurements, offered high TPR, i.e., high sensitivity, under Scenario 1-3 for Model 1-3: the approach using baseline measurements always gave TPR as high as 100% and never smaller than 98.5%; the approach using peak measurements always gave TPR as high as 100% and never smaller than 98%; our proposed approach using FPC scores gave comparable TPR over 98.5% for all first



Fig. 1 Pearson correlations between biomarkers' baseline and peak measurements, stratified by sex and masked by p-value under an α -level of 0.05

FPCs and relatively high TPR greater than 87% for the second and third FPCs (Table 3).

The FPCA-SGL approach gave relatively low FPR, as low as 0% in Scenario 2 in the case that the biomarker group with high within-group correlation was associated with the survival outcome, and no higher than 12% in the null case of Scenario 4. Notably, in every scenario under every model, this approach consistently showed smaller FPR than the approaches using baseline or peak measurements, especially under Scenario 1-2 for Model 2 (FPR ranging from 17% to 22% using baseline or peak measurements but as low as 0 - 0.5% using FPC scores). This demonstrated that our proposed two-stage FPCA-SGL approach gave much higher specificity than the simpler methods using baseline or peak measurements (Table 3).

The two comparator methods, especially the one using peak measurements, suffered from high FPR, which was particularly high in Scenario 4: the approach using peak measurements yielded an FPR of 17 - 18% under

Model 1, 13.5 - 18% under Model 2, and 40.5 - 41.5% under Model 3; the approach using baseline measurements yielded an FPR of 14 - 16% under Model 1, 15.5 - 17.5% under Model 2, and 16.0 - 17.5% under Model 3 (Table 3). This again illustrated that these two simpler approaches using baseline or peak measurements suffered from low specificity.

To investigate further the inflated FPR in Scenario 4, we considered an additional Scenario 5, a complete null case in which neither of the two biomarker groups was associated with mortality ($\alpha_1 = \alpha_2 = \alpha_3 = \alpha_4 = 0$) and observed biomarker trajectories were not censored by death times (Table 2). Simulation results showed that the FPR decreased slightly in this scenario, most notably under Model 3 (from 40.5 – 41.5% in Scenario 4 to 16.5 – 20% in Scenario 5 with peak measurements, from 7.5 – 9% in Scenario 4 to 5 – 6.5% in Scenario 5 with FPC scores), and also under Model 1 with peak measurements (from 17 – 18% in Scenario 4 to 13.5 – 16% in Scenario 5),



Fig. 2 Estimated regression coefficients $\hat{\beta}$ from SGL models fitted with the scores of the first 3 FPCs of each biomarker as exposure variables, tiles with no border or annotated numbers indicate $\hat{\beta}$ being regularized to zero (The full names of the abbreviated biomarkers are listed at the end of the manuscript)

as well as under Model 2 with FPC scores (from 10 - 12%in Scenario 4 to 4 - 5% in Scenario 5) (Supplementary Table 2). For the approach using baseline measurements, this additional Scenario 5 did not alter the baseline values thus the FPR remained similar (Supplementary Table 2).

Discussion

Our proposed FPCA-SGL approach revealed associations between several biomarker trajectories and 30-day mortality among hospitalized SARS-CoV-2 patients. In particular, renal and inflammatory biomarkers were strongly associated with mortality risks. Several studies have examined incidence of acute kidney injury (AKI) among SARS-CoV-2 patients and discovered that AKI was related to more severe outcomes including death, respiratory failure, and disseminated intravascular coagulation [7, 10, 29]. Elevated blood urea nitrogen and creatinine, as well as lower estimated glomerular filtration rate were all markers of AKI and were reported to be correlated with worse outcomes [2, 5, 7, 10, 13, 29]. Studies have indicated excessive inflammatory response as a contributory factor to SARS-CoV-2 disease severity [30]. Lymphocytes are crucial in modulating inflammatory response and maintaining immune homeostasis during viral infection [31], and research reported elevated white blood cell count and lymphopenia (low absolute



Fig. 3 Estimated regression coefficients $\hat{\beta}$ from SGL models fitted with the baseline or peak measurement of each biomarker as exposure variables, tiles with no border or annotated numbers indicate $\hat{\beta}$ being regularized to zero (The full names of the abbreviated biomarkers are listed at the end of the manuscript)

lymphocyte count) among severe SARS-CoV-2 patients [3, 4, 7]. Elevated c-reactive protein levels were also closely related to inflammation and shown to be highly associated with disease severity [32].

A limited number of studies have specifically investigated the effect of sex on the associations between biomarker levels and disease severity [12–14]. We had a relatively large cohort of 12,941 patients, thus we conducted our analyses under stratification by sex so as to better explore any potential sex modification. Interestingly, we observed associations of hematological biomarkers (hemoglobin, hematocrit, and platelets) with 30-day mortality risks only among males. As males usually experienced more severe symptoms and worse survival outcomes during SARS-CoV-2 infection [12, 14], our results may lend insight into the sex difference behind the cellular and molecular pathways underlying SARS-CoV-2 disease progression.

Methodologically, our proposed FPCA-SGL approach is an easy-to-implement and computationally efficient analytic strategy that is able to simultaneously consider multiple biomarkers as well as their longitudinal trajectories in evaluating associations with severe SARS-CoV-2 outcomes. It is a versatile alternative to existing

Table 3 Results of simulation studies

	Scenario 1		Scenario 2		Scenario 3	Scenario 4
	TPR	FPR	TPR	FPR	TPR	FPR
A. Model 1 (LME with a linear tir	me trend)					
A.1 Using baseline measur	es					
Group 1						
Biomarker 1	100.0%	N/A	N/A	10.0%	100.0%	14.0%
Biomarker 2	100.0%	N/A	N/A	10.0%	100.0%	14.5%
Group 2						
Biomarker 3	N/A	6.5%	100.0%	N/A	100.0%	15.0%
Biomarker 4	N/A	6.5%	100.0%	N/A	100.0%	16.0%
A.2 Using peak measures						
Group 1						
Biomarker 1	100.0%	N/A	N/A	5.0%	100.0%	18.0%
Biomarker 2	100.0%	N/A	N/A	5.0%	100.0%	17.5%
Group 2						
Biomarker 3	N/A	12.0%	100.0%	N/A	100.0%	17.0%
Biomarker 4	N/A	12.0%	100.0%	N/A	100.0%	17.0%
A.3 Using FPC scores						
Group 1						
Biomarker 1 FPC1	100.0%	N/A	N/A	0.0%	100.0%	9.0%
Biomarker 1 FPC2	94.0%	N/A	N/A	0.0%	92.0%	9.0%
Biomarker 1 FPC3	90.5%	N/A	N/A	0.0%	92.5%	9.0%
Biomarker 2 FPC1	100.0%	N/A	N/A	0.0%	100.0%	8.5%
Biomarker 2 FPC2	93.5%	N/A	N/A	0.0%	93.5%	9.0%
Biomarker 2 FPC3	90.5%	N/A	N/A	0.0%	88.0%	9.0%
Group 2						
Biomarker 3 FPC1	N/A	3.5%	100.0%	N/A	100.0%	5.5%
Biomarker 3 FPC2	N/A	3.0%	92.5%	N/A	91.5%	5.5%
Biomarker 3 FPC3	N/A	3.5%	91.0%	N/A	93.0%	5.5%
Biomarker 4 FPC1	N/A	3.5%	100.0%	N/A	100.0%	5.5%
Biomarker 4 FPC2	N/A	3.5%	89.5%	N/A	93.0%	5.5%
Biomarker 4 FPC3	N/A	3.5%	91.5%	N/A	87.0%	5.5%
B. Model 2 (LME with a quadrati	ic term for time)					
B.1 Using baseline measure	es					
Group 1						
Biomarker 1	100.0%	N/A	N/A	21.0%	99.0%	15.5%
Biomarker 2	100.0%	N/A	N/A	21.0%	99.0%	15.5%
Group 2						
Biomarker 3	N/A	18.0%	100.0%	N/A	100.0%	17.0%
Biomarker 4	N/A	18.0%	100.0%	N/A	100.0%	17.5%
B.2 Using peak measures						
Group 1						
Biomarker 1	100.0%	N/A	N/A	22.0%	98.0%	14.0%
Biomarker 2	100.0%	N/A	N/A	22.0%	98.0%	13.5%
Group 2						
Biomarker 3	N/A	17.0%	100.0%	N/A	100.0%	17.5%
Biomarker 4	N/A	17.0%	100.0%	N/A	100.0%	18.0%
B.3 Using FPC scores						
Group 1						
Biomarker 1 FPC1	100.0%	N/A	N/A	0.0%	98.5%	12.0%

Table 3 (continued)

	Scenario 1		Scenario 2		Scenario 3	Scenario 4
	TPR	FPR	TPR	FPR	TPR	FPR
Biomarker 1 FPC2	98.0%	N/A	N/A	0.0%	95.0%	12.0%
Biomarker 1 FPC3	90.0%	N/A	N/A	0.0%	87.5%	11.5%
Biomarker 2 FPC1	100.0%	N/A	N/A	0.0%	98.5%	12.0%
Biomarker 2 FPC2	96.0%	N/A	N/A	0.0%	89.5%	12.0%
Biomarker 2 FPC3	87.5%	N/A	N/A	0.0%	87.0%	11.5%
Group 2						
Biomarker 3 FPC1	N/A	0.5%	100.0%	N/A	100.0%	10.5%
Biomarker 3 FPC2	N/A	0.5%	97.0%	N/A	95.0%	10.5%
Biomarker 3 FPC3	N/A	0.5%	90.0%	N/A	90.0%	10.5%
Biomarker 4 FPC1	N/A	0.5%	100.0%	N/A	100.0%	10.5%
Biomarker 4 FPC2	N/A	0.5%	97.0%	N/A	90.5%	10.0%
Biomarker 4 FPC3	N/A	0.5%	89.0%	N/A	92.5%	10.5%
C. Model 3 (LME with a 3-knot s	pline function for tir	ne)				
C.1 Using baseline measure	- 2S					
Group 1						
Biomarker 1	100.0%	N/A	N/A	5.5%	98.5%	16.5%
Biomarker 2	100.0%	N/A	N/A	5.5%	98.5%	17.5%
Group 2						
Biomarker 3	N/A	5.5%	100.0%	N/A	100.0%	16.0%
Biomarker 4	N/A	5.5%	100.0%	N/A	100.0%	16.0%
C.2 Using peak measures						
Group 1						
Biomarker 1	100.0%	N/A	N/A	5.0%	99.5%	41.5%
Biomarker 2	100.0%	N/A	N/A	5.0%	99.5%	40.5%
Group 2						
Biomarker 3	N/A	10.0%	100.0%	N/A	100.0%	40.5%
Biomarker 4	N/A	10.0%	100.0%	N/A	100.0%	40.5%
C.3 Using FPC scores						
Group 1						
Biomarker 1 FPC1	100.0%	N/A	N/A	0.0%	99.0%	9.0%
Biomarker 1 FPC2	95.0%	N/A	N/A	0.0%	89.5%	8.5%
Biomarker 1 FPC3	91.0%	N/A	N/A	0.0%	89.0%	8.5%
Biomarker 2 FPC1	100.0%	N/A	N/A	0.0%	99.0%	8.5%
Biomarker 2 FPC2	94.0%	N/A	N/A	0.0%	93.5%	8.5%
Biomarker 2 FPC3	95.5%	N/A	N/A	0.0%	90.5%	8.5%
Group 2						
Biomarker 3 FPC1	N/A	3.5%	100.0%	N/A	100.0%	8.0%
Biomarker 3 FPC2	N/A	4.0%	94.5%	N/A	93.5%	8.0%
Biomarker 3 FPC3	N/A	4.0%	88.5%	N/A	88.5%	8.0%
Biomarker 4 FPC1	N/A	4.0%	100.0%	N/A	100.0%	8.0%
Biomarker 4 FPC2	N/A	4.0%	96.5%	N/A	92.5%	7.5%
Biomarker 4 FPC3	N/A	4.0%	88.0%	N/A	88.5%	7.5%

True positive rate (TPR) calculated as the proportion of simulations where truly non-zero coefficients were selected, or false positive rate (FPR) calculated as the proportion of simulations where truly zero coefficients were selected

methods concerning multiple longitudinal measurements and a survival outcome and could be applied in other areas. Using simulation studies, we demonstrated that FPCA-SGL retained high TPR and outperformed alternative approaches using baseline or peak values with respect to FPR. In particular, we observed a substantial

"survival bias" (inflated FPR) in our simulations when using peak measurements, because they are endogenous covariates, meaning their values and future paths are directly affected by the survival outcome of interest [33]. For example, if a certain biomarker has a monotonically increasing trajectory during hospitalization, the peak value observed will be higher for patients surviving longer, causing spurious associations between lower peak biomarker values and higher mortality risks. This "survival bias" resulted in high FPR in our simulation study using the peak measurement approach (Scenario 4 in Table 3) and was moderately alleviated when we did not censor the simulated biomarker trajectories based on simulated death times (Scenario 5 in Supplementary Table 2). Our FPCA-SGL approach mitigated this "survival bias" (lower FPR in Scenario 4 in Table 3) because FPCA naturally imputed biomarkers' unobserved future trajectories even after patients' deaths. Nonetheless, we did still observe some false positives using this proposed approach with FPC scores (largest FPR as 12% in Scenario 4 in Table 3).

Conclusions

We presented a two-stage analytic approach that combined FPCA and SGL to study the associations between hospitalized SARS-CoV-2 patients' multiple biomarker trajectories with their 30-day mortality rates. We demonstrated that this method had high TPR and outperformed simpler comparator approaches using biomarkers' baseline or peak measurements with respect to FPR. Using data from a retrospective cohort of 12,941 patients, we showed that renal biomarkers (blood urea nitrogen, creatinine, and estimated glomerular filtration rate), inflammatory biomarkers (c-reactive protein, white blood cell count, and absolute lymphocyte count), cardio-thrombotic biomarkers (d-dimer) were associated with 30-day mortality rates among hospitalized SARS-CoV-2 patients. Our sexstratified analysis also revealed that hematological biomarkers (hemoglobin, hematocrit, and platelets) were associated with higher mortality only among males. This study recognized the prognostic value of biomarkers as well as the underlying potential sex difference. These results provide insights into assessment of SARS-CoV-2 disease severity and effective risk stratification.

Abbreviations

- bun Blood urea nitrogen
- cre Creatinine
- gfr Estimated glomerular filtration rate
- cpk Creatine phosphokinase
- crp C-reactive protein

- wbc White blood cell count alc Absolute lymphocyte count hgb Hemoglobin hct Hematocrit Alanine aminotransferase alt Aspartate aminotransferase ast alkp Alkaline phosphatase tbili Total bilirubin
- alb Albumin
- Idh Lactate dehydrogenase
- anion Anion gap

Supplementary Information

The online version contains supplementary material available at https://doi.org/10.1186/s12874-023-02076-3.

Additional file 1. Supplementary Figures and Tables. Additional file 2. Supplementary Methods.

Acknowledgements

None.

Authors' contributions

TC conceived the idea of the study. TC and AF designed the study. All authors participated in the development of the methods. TC performed the data analysis, interpreted the findings and drafted the manuscript. HR and AF interpreted the findings and revised the manuscript. All authors read and approved the final manuscript.

Authors' informations

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Availability of data and materials

The data is not publicly available. The code for the simulation studies (both data simulation and the application of our proposed FPCA-SGL method) is available in the Github repository, https://github.com/Aimeessn/FPCA-SGL.

Declarations

Ethics approval and consent to participate

The Partners HealthCare Institutional Review Board (IRB) (#2020P000829) approved this study, including methods, the collection of curated data based on comprehensive manual chart reviews, and data extractions from electronic health records (EHR) on patients who receive care through the Mass General Brigham (MGB, formerly Partners) system. All methods were carried out in accordance with relevant guidelines and regulations. All participants provided written informed consent to participate in research.

Consent for publication

Not applicable.

Competing interests

The authors declare no competing interests.

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